

## THROMBOMODULIN DERIVATIVES AND CONJUGATES

5

### CROSS-REFERENCES TO RELATED APPLICATIONS

[0001] This application claims the benefit of U. S. Provisional Application No. 60/546,436 by Chaikof et al. filed February 20, 2004, which is incorporated herein by reference in entirety.

### STATEMENT ON FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

10

[0002] This invention was made, at least in part, with government support under Grant No. NIH RO1HL56819 awarded by the National Institutes of Health. The government has certain rights in the invention.

### BACKGROUND OF THE INVENTION

15

[0003] The transmembrane human protein thrombomodulin (TM), as a critical regulator of the protein C pathway, represents the major anticoagulant mechanism that is operative in both normal and injured blood vessels under physiologic conditions in vivo. An effective blood-contacting surface is dependent upon the presence of physiologically relevant antithrombogenic mechanisms that are incorporated into an engineered blood-material interface. Full length native human

20

TM can be incorporated into membrane-mimetic films or surfaces by fusion/adsorption processes; a major drawback of these materials, however, is a loss of TM stability and/or functional activity with time. Moreover, in existing protocols for covalent immobilization of TM onto polymeric surfaces, the protein immobilization procedure involves freely available amino or carboxyl functionalities of TM, some of which may be within or near a bioactive site. The use of such functionalities can significantly reduce the functional bioactivity of TM after surface coupling. There is therefore a need for compositions and methods in this field to serve as effective alternatives.

25

## SUMMARY OF THE INVENTION

[0004] In general the terms and phrases used herein have their art-recognized meaning, which can be found by reference to standard texts, journal references and contexts known to those skilled in the art. The following definitions are provided to clarify their specific use in the context of the invention.

[0005] When used herein, "thrombomodulin" refers to a protein molecule that is capable of involvement in the conversion of protein C to the activated protein C (protein Ca). In a particular example, the molecule relates to a specific endothelial cell receptor that forms a 1:1 stoichiometric complex with thrombin. In a preferred embodiment, the molecule is human and is also known as fetomodulin and CD141 antigen. In a particular embodiment, the term refers to a native molecule with nucleic acid or protein sequence information corresponding to that of accession number NM\_000361, Version: NM\_000361.2 GI:40288292; or Swiss-Prot: P07204.

[0006] When used herein, "derivative" refers to a variation or analog, or modification thereof, relative to a reference material. For example, a thrombomodulin derivative can refer to a mutant protein, truncation of a native protein sequence, or synthetically modified variant including modification by pegylation or conjugation such as to a polymer or surface. A derivative may employ the use of natural amino acids, non-natural amino acids, and/or other chemical moieties; whether covalently or non-covalently associated; and whether associated during translation, post-translation, or apart from translation such as in a synthetic approach; as disclosed herein or as would be understood in the art. In a particular embodiment, a derivative is a truncated, mutated, pegylated, conjugated thrombomodulin.

[0007] The following abbreviations are applicable. TM, thrombomodulin; PEG, polyethylene glycol.

[0008] It is recognized that regardless of the ultimate correctness of any mechanistic explanation or hypothesis, an embodiment of the invention can nonetheless be operative and useful.

[0009] In an embodiment, the present invention relates to the generation of novel thrombomodulin ("TM") conjugates, including soluble conjugates.

[00010] In an embodiment, the invention provides a method of site-specific PEGylation for a protein molecule. In a preferred embodiment, the protein molecule  
5 is bioactive thrombomodulin or a derivative thereof.

[00011] In an embodiment, a TM analog sequence comprises a catalytically active site capable of activating protein C (EGF 4-6 domains) and single or multiple non-natural amino acids. In a particular embodiment, TM analogs are conjugated to linear or branched natural or synthetic polymers via the non-natural amino acids. In  
10 an embodiment, the invention provides methods for conjugation of the TM conjugates to the surfaces of synthetic or natural materials, to targeting groups for site specific delivery of the agent, and/or to compounds that contain one or more additional anti-inflammatory/anti-thrombotic properties.

[00012] In an embodiment, a TM conjugate can be useful as a systemic agent for  
15 treatment of one or more conditions such as micro or macrovascular blood clots, stroke, heart attack, disseminated intravascular coagulation or other inflammatory or prothrombotic condition.

[00013] In an embodiment, the invention provides a coating of a surface of a medically implanted or human tissue or fluid contacting device. For example,  
20 implants or devices can include but are not restricted to: vascular grafts, stents, heart valves, dialysis membranes, membrane oxygenators, catheters, and guide wires to alter surface properties. A further embodiment of the invention provides a coating for living cells or tissues, including, but not restricted to smooth muscle cells, fibroblasts, endothelial cells, stem cells, chondrocytes, osteoblasts, pancreatic islets,  
25 or genetically engineered cells to establish or enhance the anti-inflammatory properties of the cells.

[00014] In an embodiment, the invention provides a covalent conjugation of truncated TM derivatives onto a blood or tissue contacting surface using natural or synthetic polymers as a spacer.

[00015] In an embodiment, the invention provides a novel method for covalent conjugation of TM to synthetic or natural materials site-specifically without loss of protein bioactivity or with substantial retention thereof. In a particular embodiment, the invention provides a relatively short recombinant TM construct containing EGF-like domains 4-6 and single or multiple non-natural amino acids optionally or preferably at the C-terminal portion of the construct. In a particular embodiment, the invention provides a modified TM through reaction with a suitable polymer spacer via one or more non-natural amino acids. In a particular embodiment, a TM is modified for further immobilization onto surfaces or for conjugation to linear or multifunctional natural or synthetic compounds that contain other anti-inflammatory or anti-thrombotic properties. In an embodiment, a bioconjugation reaction occurs in mild conditions to preserve TM bioactivity. In a preferred embodiment, polyethylene glycol ("PEG") is used as the polymer spacer.

[00016] In an embodiment, a modified construct is adapted for further immobilization onto a surface or for conjugation to linear or multifunctional natural or synthetic compounds that are capable of an anti-inflammatory or anti-thrombotic activity.

[00017] In an embodiment, PEGylation of a protein can confer one or more advantages, e.g. an increase in plasma half-life, stability against proteolytic cleavage, and a decrease of protein immunogenicity.

[00018] In an embodiment, the invention provides compositions including nucleic acid and protein molecules. In a particular embodiment, the compositions relate to sequences of the Table S1 below.

[00019] Table S1 of selected sequence listing information.

SEQ ID NO:	Brief Description	Type
1	TMb	DNA/RNA
2	automatic translation	PRT
3	TMb	PRT
4	human TM	PRT
5	PCR primer	DNA
6	PCR primer	DNA

[00020] In an embodiment, the invention provides methods and compositions relating to generation of thrombomodulin constructs comprising a non-natural amino acid. In an embodiment, a construct is thrombomodulin or a thrombomodulin derivative. In an embodiment, the invention provides recombinant expression or  
5 synthetic production of such constructs. In a preferred embodiment, the construct is generated by recombinant expression. In a preferred embodiment, the invention provides methods and compositions comprising an extracellular portion of thrombomodulin. In a preferred embodiment, the extracellular portion of thrombomodulin further comprises catalytically active sites. In a preferred  
10 embodiment, the extracellular portion of thrombomodulin is capable of activating protein C.

[00021] In a preferred embodiment, a thrombomodulin derivative comprises a single non-natural amino acid or multiple non-natural amino acids. In an embodiment, a non-natural amino acid can include those as would be understood in  
15 the art. For example, non-natural amino acids can include: methionine analogues, alanine analogues, phenylalanine analogues, leucine analogues, proline analogues and isoleucine analogues. An example of methionine analogues includes: L-2-amino-4-azido-butanoic acid.

[00022] In a preferred embodiment, a thrombomodulin derivative comprises a  
20 single non-natural amino acid at the C-terminal portion of the construct.

[00023] In an embodiment, the invention provides a thrombomodulin construct wherein the construct is conjugated to a natural or synthetic polymer or other natural molecule such as an antibody or other ligand recognition molecule. In an  
embodiment, the conjugation is via at least one non-natural amino acid in the  
25 recombinant protein.

[00024] In an embodiment, a synthetic polymer for conjugation to a construct can include a linear or branched synthetic polymer. For example, a linear or branched synthetic polymer can include: poly(t-butyl acrylate), poly(t-butyl methacrylate), polyacrylamide, glycolipid and their mimetics; and other polymers as would be  
30 understood in the art. Examples of a natural polymer include: glycoproteins and their mimetics, poly(arginine), polysaccharides and their mimetics; and other

polymers as would be understood in the art. In an embodiment, a ligand recognition molecule is antifibrin antibody.

[00025] In an embodiment, a construct is conjugated to a linear or branched poly(ethylene glycol) molecule. In a preferred embodiment, the construct is  
5 conjugated to linear poly(ethylene glycol).

[00026] In an embodiment, the invention provides a thrombomodulin construct conjugated to a natural or synthetic polymer for surface anchoring. In an embodiment, the natural or synthetic polymer is multifunctional. In a preferred  
10 embodiment, the construct is conjugated to poly(ethylene glycol) for surface anchoring of the conjugate. In an example, an anchoring group includes: biotin, conjugated diene, azide, alkyne, diphenylphosphine, triarylphosphine; and other groups as would be understood in the art. In an example, a surface targeting group  
15 includes: sialyl-Lewis X; an antibody, Fab fragment or the like, or other analogous protein or non-protein recognition molecule (including an aptamer) capable of recognizing VCAM-1, ICAM-1, or other inflammatory cell surface proteins; antifibrin antibody; streptavidin, azide, alkyne, N-( $\epsilon$ -maleimidocaproyl; and others as would be understood in the art.

[00027] In an embodiment, a construct is conjugated to a synthetic polymer for anchoring to a surface of a synthetic material or a natural material. In an example,  
20 synthetic materials include: poly(tetrafluoroethylene) , polysiloxanes, poly(ether urethane urea), poly(lactic acid-co-glycolic acid), a glass surface and derivatives; and other materials as would be understood in the art. In an example, natural materials include: cells, tissues, and blood vessels.

[00028] In an embodiment, the invention provides compositions and methods of a  
25 surface coating for a medically implanted or human tissue or fluid contacting device including but not restricted to vascular grafts, stents, heart valves, dialysis membranes, membrane oxygenators, catheters, or guide wires. In an embodiment, the surface coating alters a surface property of the implant or the device.

[00029] In an embodiment, the invention provides compositions and methods for  
30 coating a surface of living cells or tissues, including, but not restricted to smooth

muscle cells, fibroblasts, endothelial cells, stem cells, chondrocytes, osteoblasts, pancreatic islets, or genetically engineered cells.

[00030] In an embodiment, the invention provides a recombinant thrombomodulin construct conjugated to a multifunctional natural or synthetic polymer, wherein the polymer is capable of an anti-inflammatory or anti-thrombotic property. In an embodiment, the construct is conjugated to a synthetic polymer comprising one or more anti-inflammatory groups. In an embodiment, the synthetic polymer comprises one or more additional anti-inflammatory groups. In an example, an anti-inflammatory group includes sialic acids and their mimetics/derivatives; and other groups as would be understood in the art.

[00031] In an embodiment, the invention provides a construct conjugated to a synthetic polymer further comprising an anti-coagulant or anti-thrombotic group. In an example, an anti-coagulant or anti-thrombotic group includes heparin and its mimetics/derivatives; and other groups as would be understood in the art.

[00032] In an embodiment, the invention provides a systemic agent for treatment of a medical condition, wherein the condition relates to a microvascular or macrovascular blood clot, stroke, heart attack, disseminated intravascular coagulation, or other inflammatory or prothrombotic condition. In an embodiment, the invention provides a method of treatment of the medical condition by administering to a patient in need a construct of the invention.

[00033] In an embodiment, the invention provides a truncated thrombomodulin protein derivative comprising EGF (4-6) like domains, a substitution of Leucine for methionine at position 388, and a GGM amino acid motif appended at a carboxy terminus of said derivative. In an embodiment, the GGM protein motif is expressed as a protein motif with a non-natural amino acid corresponding to the M amino acid residue. In an embodiment, the invention provides SEQ ID NO:3.

[00034] In an embodiment, the invention provides a truncated thrombomodulin derivative conjugate comprising a truncated thrombomodulin derivative and a polymer; wherein the thrombomodulin derivative comprises EGF (4-6) like domains, a substitution of Leucine for methionine at position 388, and a GGM amino acid motif

appended at a carboxy terminus of said derivative. In an embodiment, the polymer comprises polyethylene glycol.

[00035] In an embodiment, the invention provides a truncated thrombomodulin nucleic acid derivative comprising EGF (4-6) like domains, a substitution of Leucine for methionine at position 388, and a nucleic acid sequence capable of encoding a Gly Gly Met motif appended at a carboxy terminus of said derivative. In an embodiment, the nucleic acid sequence comprises SEQ ID NO:1.

[00036] In an embodiment, the invention provides a method of generating a purified truncated thrombomodulin derivative protein, wherein the protein comprises EGF (4-6) like domains, a substitution of Leucine for methionine at position 388, and a non-natural amino acid; comprising the steps of providing a truncated thrombomodulin nucleic acid sequence; recombinantly expressing said nucleic acid sequence in the presence of a non-natural amino acid precursor; and purifying a recombinant expression product; thereby generating a purified truncated thrombomodulin derivative protein. In an embodiment, the nucleic acid sequence is SEQ ID NO:1.

## BRIEF DESCRIPTION OF THE FIGURES

[00037] Scheme A. Synthesis of the methyl-PEG-triarylphosphine conjugate **2** and its conjugation to azido-functionalized TMB.

[00038] Figure 1 is a schematic structure of the targeted truncated TM, whose amino-acid sequence is encoded by the gene TM<sub>GGM</sub>.

[00039] Figure 2 illustrates results of Western blot analysis of TM derivatives (4-20% SDS-PAGE gel): (1) purified **TM<sub>A</sub>**; (2) enterokinase cleavage of **TM<sub>A</sub>** leading to the target protein **TM<sub>B</sub>**; (3-5) conjugation of **2** to **TM<sub>B</sub>** over time: 4h, 8h and 24h, respectively. M: Molecular weight marker proteins.

[00040] Figure 3 illustrates MALDI-TOF Mass Spectra of (a) the starting polymer methyl-PEG-amine (average molecular weight detected: 5,668 D) and (b) the



methyl-PEG-triarylphosphine conjugate **2** (average molecular weight detected: 6,034 D, calculated 6014 D).

[00041] Figure 4 illustrates (a) Western blot analysis; and (b) barium chloride/iodine staining of a 10% SDS-PAGE gel: (1) and (3) reaction mixture of the  
5 bioconjugation of **2** to **TM<sub>B</sub>** after 36h; (2) initial azido-functionalized **TM<sub>B</sub>**. M: Molecular weight marker proteins.

[00042] Figure 5 illustrates a genetically directed synthesis using non-natural amino acids and a conjugation reaction.

[00043] Figure 6 illustrates bioorthogonal surface ligation of azidolactose (A) and  
10 azidoTM (B) via azide-alkyne [3+2] cycloaddition ("Click" chemistry). Surface bound carbohydrates and TM were visualized by staining with a FITC-lectin (A) and FITC anti-S-tag MAb (B), respectively.

#### DETAILED DESCRIPTION OF THE INVENTION

[00044] The invention may be further understood by the following non-limiting  
15 examples.

##### **[00045] EXAMPLE 1. C-Terminal site-specific PEGylation of a truncated thrombomodulin with full bioactivity**

[00046] Abstract: Addition of polyethylene glycol to bioactive proteins (PEGylation) can improve their plasma half-life, enhance stability against proteolytic cleavage, and  
20 may also decrease protein immunogenicity. PEGylation can involve reaction to available lysine amino groups, some of which may be within or near a bioactive site. Often PEGylation protocols are nonspecific and result in a loss of protein activity. We disclose a strategy for site-specific PEGylation of a thrombomodulin derivative (TM) at the C-terminus. A truncated TM mutant consisting of epidermal growth  
25 factor (EGF)-like domains 4-6 was expressed using recombinant techniques in *Escherichia coli* with a C-terminal azido-methionine. The TM mutant was site-specifically conjugated to a methyl-PEG-triarylphosphine compound via Staudinger reaction. Enzymatic activity of the TM construct before and after PEGylation was

substantially similar, which confirms the utility of this site-specific PEGylation method and molecules thereby produced.

[00047] Introduction: Pegylation of proteins increases both their molecular size and steric hindrance, which can result in an increase of protein plasma half-life and resistance to proteolytic cleavage. In addition, protein immunogenicity may be decreased (1,2). Characteristically, PEGylation usually involves reaction to available lysine amino groups, some of which may be within or near a bioactive site. Thus, protocols are often nonspecific and result in a loss of protein activity (3,4). For example, Han *et al.* (5) conjugated TM to PEG via trichlorotriazine as a coupling agent for immobilization onto a glass surface. Despite successful immobilization, reduced TM activity was noted presumably due to alteration of protein conformation after PEGylation. To overcome the limitations of current conjugation strategies, several approaches have been proposed. Site-specific PEGylation can be achieved through the introduction of a cysteine residue in engineered proteins with a free thiol available for conjugation reactions (6,7). As a consequence, several PEG derivatives have been developed for that purpose (8,9,10,11). The efficacy of this approach is compromised, however, by a low yield of PEGylated protein and often by a substantial loss of activity, for example if abnormal protein folding is induced by the introduction of the cysteine residue (12). As an alternative strategy, Yamamoto *et al.* (13) reported site-specific PEGylation of a lysine-deficient tumor necrosis factor- $\alpha$  at its N-terminus.

[00048] We report herein the novel use of a strategy for site-specific PEGylation of human thrombomodulin. This transmembrane protein is a critical regulator of the protein C pathway and represents a major anticoagulant mechanism that is operative under physiologic conditions *in vivo* (14,15). TM is a cofactor for thrombin-catalyzed activation of protein C, enhancing the rate of the reaction by 1000-fold (16). In order to closely mimic the TM structure as it appears at the cell surface (17,18), and consequently preserve its bioactivity, we investigated the PEGylation at the C terminus. Using a genetically-directed synthesis in *Escherichia coli*, we first expressed a short TM construct containing EGF-like domains 4-6 and an azido-functionalized methionine analog (19) as a C-terminal linker. The PEGylation was

then achieved through Staudinger ligation (20) with a suitably engineered PEG derivative.

#### [00049] EXPERIMENTAL PROCEDURES

[00050] Materials. All chemical reagents were obtained from Sigma Chemical Corporation (St. Louis, MS). The methyl-PEG-amine 5,000 was purchased from Netkar Corp. (Huntsville, AL). The BamHI and Shrimp Alkaline Phosphatase enzymes were obtained from New England Biolabs, Inc. (Beverly, MA). The Quikchange Site-Directed Mutagenesis kit was from Stratagene (La Jolla, CA). E.coli strain B834 (DE3), plasmid pET-39b(+), S-Tag Rapid Assay kit and Site-Specific Enterokinase Cleavage and Capture Kits were from Novagen (Madison, WI). All plasmid purification kits were purchased from QIAGEN Inc. (Chatsworth, CA). The mouse monoclonal antibody to human thrombomodulin was from COVANCE Corp. (Richmond, CA). Synthetic oligonucleotides were purchased from Integrated DNA Technologies, Inc. (Coralville, IA). Purified recombinant human PC and human  $\alpha$  thrombin were from Haematologic Technologies Inc. (Essex Junction, VT). Human anti-thrombin, recombinant human Thrombomodulin (soluble truncated form of thrombomodulin that lacks the putative transmembrane and the cytoplasmic domains, approximate weight 68 kD) and chromogenic substrate SPECTROZYME PCa were purchased from American Diagnostica Inc. (Stamford, CT). All reagents for manipulating DNA and bacteria were sterilized by autoclave.

[00051] Instrumentation. MALDI-TOF mass spectrometry data were performed on an Applied Biosystem Voyager-DE<sup>TM</sup> STR Biospectrometry<sup>TM</sup> Workstation MALDI-TOF Mass Spectrometer using an 2-(4-hydroxy-phenylazo)benzoic acid matrix. <sup>1</sup>H, <sup>13</sup>C and <sup>31</sup>P NMR spectra were recorded at 600 MHz (<sup>1</sup>H, <sup>13</sup>C) and 242 MHz (<sup>31</sup>P) on a Varian INOVA in CDCl<sub>3</sub> (internal Me<sub>4</sub>Si,  $\delta$  (delta) ppm). Optical density was recorded on a Varian Cary 50 Bio UV-visible spectrophotometer.

[00052] Synthetic Gene Construction. A DNA fragment encoding for EGF (4-6) domains of human TM was obtained by polymerase chain reaction using primers 5'-TACCCTAACTACGACCTGGTG-3' (SEQ ID NO:5) and 5'-TATGAGCAAGCCCGAATG-3' (SEQ ID NO:6). Through a series of intermediate constructs, this fragment was used to generate a gene containing a Leucine (Leu)

substitution for Methionine-388 (Met-388), N-terminal and C-terminal BamH I sites and a C-terminal linker GlyGlyMet using site-directed mutagenesis. The final construct (TM<sub>GGM</sub>) was then inserted using the BamH I site of the expression plasmid pET-39b(+). All mutations were verified through sequence analysis.

- 5 [00053] Protein expression and purification. pET39b(+)-TM<sub>GGM</sub> was transformed into the *E. coli* methionine auxotroph B834(DE3). M9 minimal medium (500 mL) supplemented with 1 mM MgSO<sub>4</sub>, 0.4 wt% glucose, 1 mg/L thiamine chloride, 0.1 mM CaCl<sub>2</sub>, kanamycin (30 mg/L) and all proteinogenic amino acids (40mg/L) was inoculated with 20 mL of an overnight culture of the transformed cells. When the  
10 turbidity of the culture reached an OD<sub>600</sub> of 0.8, protein expression was induced by addition of isopropyl-β (beta)-D-thiogalactopyranoside (IPTG) to a final concentration of 0.5 mM. After 5min, the medium was exchanged to remove methionine, cells were sedimented (4000g, 20min), and the cell pellet washed twice with 200 mL of 1xM9 salts. Cells were resuspended in 500 mL of the M9 minimal medium  
15 described above, without methionine but supplemented with 100 mg/L of azido-functionalized methionine analog (19). A culture lacking methionine served as the negative control. Cultures were grown for 4.5 h at 37 °C.

- [00054] The expression of the TM protein was analyzed by 4-20% gradient SDS-PAGE gel electrophoresis and visualized by Western blot analysis using a mouse  
20 monoclonal antibody to human thrombomodulin. The target protein was expressed as a N-terminal Dsba enzyme fusion to a leader sequence containing an *Enterokinase* cleavage site, hexahistidine, and S-tags (protein **TM<sub>A</sub>**). S-Tag Rapid Assay was used to quantify protein concentration and therefore expression yield, which averaged 17 mg/L of cell culture. **TM<sub>A</sub>** was purified from the cell pellet by  
25 using immobilized metal-affinity chromatography on TALON resin (Clontech Laboratories, Inc.) under native conditions using an imidazole gradient for elution of the target polypeptide. The cells were first harvested by centrifugation at 4 °C at 10,000g for 30 min and resuspended in 25 mL of lysis buffer (300 mM NaCl, 50 mM NaH<sub>2</sub>PO<sub>4</sub>, 10% glycerol, 1 mg/mL lysozyme, 10 μg/mL PMSF, pH 8). After  
30 incubation on ice for 30min, the cell lysate was clarified by centrifugation at 10,000g for 20 min. The soluble extract was then loaded onto a column containing TALON metal affinity resin (25 mL), which had been preequilibrated with lysis buffer. The

weakly binding proteins were removed by rinsing the column with 125 mL wash buffer (300 mM NaCl, 50 mM NaH<sub>2</sub>PO<sub>4</sub>, 10% glycerol, 20mM imidazole, pH 8). **TM<sub>A</sub>** was eluted by the addition of 50 mL of elution buffer (300 mM NaCl, 50 mM NaH<sub>2</sub>PO<sub>4</sub>, 10% glycerol, 250mM imidazole, pH 8). The chromatographic fractions were analyzed by 4-20% gradient SDS-PAGE gel electrophoresis and visualized by Western blot analysis using mouse monoclonal antibody to human thrombomodulin. The nitrocellulose membrane was developed using the ECL plus Western blotting detection kit (Amersham Biosciences, UK). *Enterokinase* cleavage removed the fusion tag and generated the target protein (**TM<sub>B</sub>**). N-terminal sequencing, amino acid compositional and mass analysis (SELDI-TOF) confirmed the integrity of **TM<sub>B</sub>**: (Mass detected (m/z): 16,545.2 D (calculated 16,540.1 D)).

[00055] Synthesis of methyl-PEG-triarylphosphine (2). 1,3-

Dicyclohexylcarbodiimide (DCC) (5.7 mg, 27  $\mu$ mol) was added to a solution of phosphino reagent (1) (10.3 mg, 28  $\mu$ mol) in anhydrous CHCl<sub>3</sub> (2 mL) at room temperature under argon. The mixture was stirred for 1h, followed by the addition of methyl-PEG-amine (m-PEG-NH<sub>2</sub> (M<sub>n</sub> 5kD)) (120 mg, 24  $\mu$ mol) dissolved in CHCl<sub>3</sub> (2 mL). The mixture was subsequently agitated for 2.5 days under argon. The methyl-PEG-triarylphosphine conjugate was recovered by precipitation in diethyl ether (200 mL) and filtration. The precipitate was dissolved in water. The aqueous solution was filtrated and lyophilized to afford the final product with 50% of substitution. The product was used in the Staudinger reaction without further purification. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  3.38 (s, 3H, O-CH<sub>3</sub> (PEG)), 3.45-3.90 (m, 448 H, O-CH<sub>2</sub> (PEG)), 7.47-7.55 (m, 6H), 7.67-7.70 (m, 5H), 7.91-7.94 (m, 1H), 8.12-8.13 (m, 1H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  40.0, 41.3, 52.5, 59.3, 70.2, 70.6, 71.1, 72.0, 128.5, 128.7, 128.8, 130.5, 131.0, 131.3, 132.0, 133.3, 134.1, 137.7, 138.1, 166.5, 166.7. <sup>31</sup>P NMR (242 MHz, CDCl<sub>3</sub>):  $\delta$ -3.11. MALDI-TOF: 6034 D (calculated 6014 D).

[00056] **TM-PEG Conjugation.** To a solution of **TM<sub>B</sub>** in Phosphate Buffered Saline pH 7.4 (300-500 nM, 100  $\mu$ L) was added a large excess (1 mg) of lyophilized methyl-PEG-triarylphosphine and the mixture was subsequently heated at 37°C for 36 h to obtain maximum conjugation. The mixture was divided in half and two sets of reactions were subjected to SDS/PAGE in parallel. Proteins from the first gel were transferred to nitrocellulose membrane and visualized by Western blot analysis using

a mouse monoclonal antibody to human thrombomodulin. This procedure monitored the presence of TM<sub>B</sub> proteins that had reacted with the phosphine, and were therefore PEG-labeled, as well as unreacted TM<sub>B</sub>. The second gel was stained with barium chloride/iodine solutions to reveal PEGylated molecules only (21).

- 5 [00057] **TM enzymatic activity assay.** A saturated concentration of TM<sub>B</sub> in 20  $\mu$ L of assay buffer (20 mM Tris-HCl pH 7.5, 100 mM NaCl, 2.5 mM CaCl<sub>2</sub>) was incubated for 30 min at 37 °C with 10 nM human  $\alpha$  thrombin and varying amounts of human protein C (0-8  $\mu$ M). Reactions were quenched by addition of 5  $\mu$ L of human antithrombin (10.7  $\mu$ M) and incubation for 5 min at room temperature. The amount  
10 of activated protein C formed was determined by addition of 275  $\mu$ L of SPECTROZYME PCa (218  $\mu$ M), followed by incubation for 20 min at room temperature. SPECTROZYME PCa is a chromogenic substrate used for the quantification of activated protein C in solution. One mole of the substrate is hydrolyzed by activated protein C producing one mole of p-nitroaniline (pNA). The  
15 pNA concentration was determined by UV spectrophotometry at 407nm using the equation:

[00058]  $OD_{407nm} = -0.0014 + 0.0096 [pNA]$  (1)

- [00059] where equation (1) was obtained by optical density measurements of pNA solutions of known concentrations. The corresponding rate (mol/min) of p-nitroaniline formed ( $R_{pNA}$ ) was obtained by dividing the pNA concentration by the 20  
20 min assay time.

[00060] A second standard plot was used to determine the activated protein C concentration from  $R_{pNA}$ , where

[00061]  $R_{pNA} = -0.059 + 0.0671 [APC]$  (2)

- 25 [00062] To obtain equation (2), commercial activated protein C was dissolved in assay buffer at various concentrations and incubated with Spectrozyme PCa for 20 min at room temperature. The corresponding rate (mol/min) of activated protein C formed ( $R_{APC}$ ) by the TM-thrombin complex was obtained by dividing the observed APC concentration by the 30 min assay time.

[00063] Values obtained for control experiments without TM were subtracted to give final rates of activated protein C produced. Michaelis-Menten parameters ( $k_{cat}$ ,  $K_m$ ) were calculated from a plot of  $R_{APC}$  versus protein C concentration.

## [00064] RESULTS

5 [00065] Our approach utilized the three consecutive EGF-like domains 4-6 (EGF4-6) of human TM. We analyzed that the corresponding amino-acid sequence contains only one methionine residue (Met-388) and considered its possible impact on protein bioactivity. We also noted that the mutation of Met-388 to leucine (Leu) could contribute resistance to oxidative inactivation while allowing the potential for  
10 greater enzymatic activity than the native TM protein. We therefore have made the significant finding that a truncated TM fragment containing EGF domains 4-6 with the insertion of a C-terminal non-natural methionine analog can provide a good target for site-specific PEGylation.

[00066] To create a short recombinant bioactive TM mutant, we first used the DNA  
15 fragment encoding for the amino-acids sequence 349 to 492 to generate a gene containing a Met-388-Leu substitution and a C-terminal linker GlyGlyMet using site-directed mutagenesis (**Figure 1**). The final construct ( $TM_{GGM}$ ) was then inserted in the expression plasmid pET-39b(+). This plasmid contains a leader gene sequence coding for the *Dsba* enzyme, which is a periplasmic enzyme catalyzing the formation  
20 and isomerization of disulfide bonds of expressed proteins. Therefore, we expected the target TM mutant to possess all disulfide bonds required for proper protein folding and enzyme activity. Expression under the induction of IPTG using the *E. coli* methionine auxotroph B834 (DE3)/pET39b(+)-***TM*<sub>GGM</sub>** in cultures depleted of methionine and supplemented with an azido-functionalized methionine analog,  
25 azidohomoalanine (19), afforded the target protein as a N-terminal *Dsba* enzyme fusion to a leader sequence containing *enterokinase* recognition sequence, hexahistidine and S-tags. Protein expression was monitored by SDS/PAGE analysis and visualized by Western blot analysis. The target protein was not observed in negative control culture, whereas TM fused to the leader sequence (***TM*<sub>A</sub>**) was clearly  
30 detected in positive control cultures supplemented with azidohomoalanine. The accumulation of ***TM*<sub>A</sub>** was taken as preliminary evidence for incorporation of the non-natural amino acid. ***TM*<sub>A</sub>** was purified from the cell pellet by using immobilized metal-

affinity chromatography with stepwise imidazole gradient elution under native conditions. *Enterokinase* cleavage removed the fusion tag and generated the target proteins (**TM<sub>B</sub>**). N-terminal sequencing, amino acid compositional and mass analysis (mass detected: 16,545D) confirmed the integrity of **TM<sub>B</sub>**. **TM<sub>B</sub>** was characterized by  
5 a band at 33kD on Western blot analysis (Figure 2), which is consistent with TM associating into dimers even under denaturing conditions (25).

[00067] Having expressed an azide-modified form of TM, we then investigated the selective Staudinger ligation of this protein with a methyl-PEG-triarylphosphine conjugate **2** (**Scheme A**). We choose this ligation protocol in order to minimize the  
10 risk of side reactions that might alter TM structure and activity. In its classical form, the Staudinger reaction meets many of the criteria required of a chemoselective ligation in a cellular environment. The phosphine and the azide react rapidly, selectively, and in high yield, in water at room temperature. We used **1** in the synthesis of methyl-PEG-triarylphosphine conjugate **2** (see Kiick *et al.* (19);  
15 preparation of a triarylphosphine compound **1** that was used for the preparation of a protein-Flag conjugate, as well as for labeling cell surface azide-bearing sialic acids with a biotinylated phosphine (20)).

[00068] The carboxylic acid group of **1** was reacted with the amino group of a commercially available derivative, methyl-PEG-amine (m-PEG-NH<sub>2</sub> (5kD)) with  
20 amide bond formation (**Scheme A**). The incorporation of a phosphino group in PEG was assessed by <sup>1</sup>H, <sup>13</sup>C, and <sup>31</sup>P NMR, as well as by MALDI-TOF spectroscopy (**Figure 3**). The average molecular weight of 6,034 D observed for the methyl-PEG-triarylphosphine conjugate **2** is consistent with an expected mass increase of 346 D, as a result of linking the phosphino compound **1** to m-PEG-NH<sub>2</sub> (average molecular  
25 weight observed: 5,668 D).

[00069] The addition of m-PEG-NH<sub>2</sub> to **1** dramatically enhances the water solubility of the hydrophobic phosphino group. A large excess of lyophilized methyl-PEG-triarylphosphine conjugate **2** (1 mg) was added to 100 μL of an aqueous solution of  
30 **TM<sub>B</sub>** in PBS at pH 7.4 and Staudinger ligation performed at 37 °C for 24h. The formation of the **TM<sub>B</sub>**-PEG conjugate was followed by Western blot analysis of the reaction mixture over time (**Figure 2**). The gel shows a band at higher molecular weight than the initial **TM<sub>B</sub>** with a simultaneous disappearance of the **TM<sub>B</sub>** band.



These results indicate that the Staudinger reaction proceeded efficiently with incorporation of the PEG polymer into the TM construct. As the reaction was not complete after 24h, we allowed it to proceed for an additional 12h. Western blot analysis (**Figure 4a**) and barium chloride/iodine staining (**Figure 4b**) of SDS-PAGE gels were performed in parallel to monitor the formation of the **TM<sub>B</sub>-PEG** conjugate. Disappearance of the band characteristic of the unconjugated **TM<sub>B</sub>** revealed that bioconjugation of **TM<sub>B</sub>** to PEG was substantially (approximately 100%) complete. The SDS gel run in parallel under identical conditions but stained with barium chloride/iodine confirmed the presence of PEG in the bioconjugate. The presence of a low molecular weight band represents unreacted methyl-PEG-triarylphosphine **2**.

[00070] Enzyme activity was assessed to determine the effect of PEGylation on TM cofactor activity (**Table 1**).

[00071] Table 1. Michaelis-Menten parameters for protein C activation by TM derivatives.

Parameter	<b>TM<sub>A</sub></b>	<b>TM<sub>B</sub></b>	<b>PEG-TM<sub>B</sub></b>	<b>Commercial TM</b>
$K_M$ ( $\mu\text{M}$ )	$0.9 \pm 0.2$	$1.0 \pm 0.5$	$1.0 \pm 0.5$	$0.7 \pm 0.1$
$k_{\text{cat}}$ ( $\text{min}^{-1}$ )	$0.22 \pm 0.05$	$0.16 \pm 0.05$	$0.20 \pm 0.05$	$0.14 \pm 0.02$
$k_{\text{cat}}/K_M$ ( $\text{min}^{-1} \cdot \mu\text{M}^{-1}$ )	$0.26 \pm 0.10$	$0.16 \pm 0.05$	$0.20 \pm 0.05$	$0.21 \pm 0.05$

[00072] We initially investigated the activity of **TM<sub>A</sub>** and **TM<sub>B</sub>** mutants, as well as a commercial recombinant human TM mutant consisting solely of the extracellular domain (American Diagnostica Inc.). Clarke et al. (23) have reported that the Met-388-Leu mutation results in a 2-fold increase in  $k_{\text{cat}}$  for the activation of protein C by a thrombin-TM fragment complex. Although we were unable to observe this enhancement of activity, Michaelis-Menten parameters were similar for **TM<sub>A</sub>** and **TM<sub>B</sub>**, as well as for the commercial human TM protein. We inferred that the generated TM mutants exhibited the requisite conformation and structure necessary for proper thrombin binding and protein C activation. Significantly, the incorporation of PEG into **TM<sub>B</sub>** did not affect cofactor activity, indicating that this site-specific PEGylation scheme did not interfere with thrombin binding and protein C activation. Parenthetically, despite a concerted effort by our group, previous attempts to

generate an enzymatically active truncated TM mutant with a C terminal cysteine, as a potential bioconjugation site, were unsuccessful using both yeast and *E. coli* expression systems. We speculate that an additional terminal cysteine likely interfered with normal protein folding dependent upon the presence of native  
5 disulfide loops in the EGF sequences.

#### [00073] CONCLUSIONS

[00074] A human thrombomodulin derivative containing EGF(4-6)-like domains designed with an azido-functionalized methionine C-terminal linker was successfully synthesized using a genetic engineering strategy. The protein exhibited bioactivity  
10 towards protein C that was comparable to native human TM. The TM construct was successfully conjugated to a novel engineered methyl-PEG-triarylphosphine compound *via* Staudinger reaction under mild conditions. Enzyme activity before and after PEGylation was substantially similar indicating the utility and successful application of the site-specific PEGylation method. To our knowledge, this report is  
15 the first to describe site-specific PEGylation of a thrombomodulin mutant with retention of substantially full bioactivity.

#### [00075] References for Example 1:

1. Veronese, F. M. (2001) Peptide and protein PEGylation: a review of problems and solutions. *Biomaterials* 22, 405-417.
- 20 2. Roberts, M. J., Bentley, M. D., and Harris, J. M. (2002) Chemistry for peptide and protein PEGylation. *Adv Drug delivery reviews* 54, 459-476.
3. Katre, N.V. (1993) The conjugation of proteins with polyethylene glycol and other polymers. *Altering properties of proteins to enhance their therapeutic potential. Adv Drug Delivery Rev* 10, 91-114.
- 25 4. Monkarsh, S. P., Ma, Y. M., Aglione, A., Bailon, P., Ciolek, D., DeBarbieri, B., Graves, M. C., Hollfelder, K., Michel, H., Palleroni, A., Porter, J. E., Russoman, E., Roy, S., and Pan, Y. C. E. (1997) Positional isomers of monopegylated interferon 2a: isolation, characterization, and biological activity. *Anal. Biochem.* 247, 434-440.
- 30 5. Han, H-S., Yang, S-L., Yeh, H-Y., Lin, J-C., Wu, H-L., and Shi, G-Y. (2001) Studies of a novel human thrombomodulin immobilized substrate: surface characterization and anticoagulation activity evaluation. *J. Biomater. Sci. Polymer edn.* 12, 1075-1089.
- 35 6. Tsutsumi, Y., Onda, M., Nagata, S., Lee, B., Kreitman, R. J., and Pastan, I. (2000) Site-specific chemical modification with polyethylene glycol of recombinant immunotoxin anti-Tac(Fv)-PE38 (LMB-2) improves antitumor activity and reduces animal toxicity and immunogenicity. *Proc. Natl. Acad. Sci. USA* 97, 8548-8553.

7. He, X. H., Shaw, P. C., Xu, L. H., and Tam, S. C. (1999) Site-directed polyethylene glycol modification of trichosanthin: effects on its biological activities, pharmacokinetics, and antigenicity. *Life Sci.* 64, 1163-1175.
- 5 8. Goodson, R. J., and Katre, N. V. (1990) Site-directed pegylation of recombinant interleukin-2 at its glycosylation site. *Bio-Technology* 8, 343-346.
9. Kogan, T. P. (1992) The synthesis of substituted methoxy-poly(ethylene glycol) derivatives suitable for selective protein modification. *Synth. Commun.* 22, 2417-2424.
- 10 10. Morpurgo, M., Veronese, F. M., Kachensky, D., and Harris, J. M. (1996) Preparation and characterization of poly(ethylene glycol) vinyl sulfone. *Bioconjug. Chem.* 7, 363-368.
11. Woghiren, C., Sharma, B., and Stein, S. (1993) Protected thiol-polyethylene glycol: a new activated polymer for reversible protein modification. *Bioconjug. Chem.* 4, 314-318.
- 15 12. Kuan, C. T., Wang, Q. C., and Pastan, I. (1994) *Pseudomonas* exotoxin A mutants. Replacement of surface-exposed residues in domain II with cysteine residues that can be modified with polyethylene glycol in a site-specific manner. *J. Biol. Chem.* 269, 7610-7616.
- 20 13. Yamamoto, Y., Tsutsumi, Y., Yoshioka, Y., Nishibata, T., Kobayashi, K., Okamoto, T., Mukai, Y., Shimizu, T., Nakagawa, S., Nagata, S., and Mayumi, T. (2003) Site-specific PEGylation of a lysine-deficient TNF- $\alpha$  with full bioactivity. *Nature Biotechnology* 21, 546-552.
14. Weiler, H., and Isermann, B. H. Thrombomodulin. (2003) *J. Thrombosis and Haemostasis* 1, 1515-1524.
- 25 15. Kalafatis, M., Egan, J. O., Van't Veer, C., Cawthorn, K. M., and Mann, K. G. (1997) The regulation of clotting factors. *Crit. Rev. Eukaryot. Gene Expr.* 7, 241-280.
16. Esmon, N. L., Owen, W.G., Esmon, C.T. (1982) Isolation of a membrane-bound cofactor for thrombin-catalyzed activation of protein-C. *J. Biol. Chem.* 257, 859-864.
- 30 17. Esmon, C. T. (1992) The regulation of clotting factors. *Arteriosclerosis and Thrombosis* 12, 135-145.
18. Sadler, J. E. Thrombomodulin structure and function. (1997) *Thrombosis and Haemostasis* 78, 392-395.
- 35 19. Kiick, K. L., Saxon, E., Tirrell, D. A., Bertozzi, C. R. (2002) Incorporation of azides into recombinant proteins for chemoselective modification by the Staudinger ligation. *PNAS* 99, 19-24.
20. Saxon, E., Bertozzi, C. R. (2000) Cell surface engineering by a modified Staudinger reaction. *Science* 287, 2007-2010.
- 40 21. Kurfurst, M. M. (1992) Detection and molecular weight determination of polyethylene glycol-modified hirudin by staining after sodium dodecyl sulfate-polyacrylamide gel electrophoresis. *Anal. Biochem.* 200, 244-248.
22. Parkinson, J. F., Nagashima, M. M, Kuhn, I., Leonard, J., Morser, J. (1992) Structure-Function studies of the epidermal growth-factor domains of human thrombomodulin. *Biochem Biophys. Res. Commun.* 185, 567-576.
- 45 23. Clarke, J. H., Light, D. R., Blasko, E. J., Parkinson, F., Nagashima, M., McLean, K., Vilander, L., Andrews, W. H., Morser, J., Glaser, C. B. (1993) The short loop between epidermal growth factor-like domain-4 and domain-5 is critical for human thrombomodulin function. *J. Biol. Chem.* 268, 6309-6315.

24. Nagashima, M., Lundh, E., Leonard, J. C., Morser, J., Parkinson J. F. (1993) Alanine-scanning mutagenesis of the epidermal growth factor-like domains of human thrombomodulin identifies critical residues for its cofactor activity. J. Biol. Chem. 268, 2888-2892.

25. Glaser, C. B., Morser, J., Clarke, J. H., Blasko, E., McLean, K., Kuhn, I., Chang, R.J., Lin, J.-H., Vilander, L., Andrews, W. H., and Light, D. R. (1992) Oxidation of a specific methionine in thrombomodulin by activated neutrophil products blocks cofactor activity. A potential rapid mechanism for modulation of coagulation. J. Clin. Invest. 90, 2565-2573.

**[00076] EXAMPLE 2. Synthesis of a recombinant thrombomodulin conjugate for immobilization onto a thin film.**

[00077] Covalent immobilization of TM onto polymeric surfaces has been investigated (AD, AE). In all cases, the conjugation scheme utilized a non-site specific carbodiimide based coupling reaction in which TM was coupled to the substrate via any freely available amino or carboxyl functionality on the protein surface. Consequently, TM bioactivity was often reduced after surface coupling.

[00078] We have demonstrated that full length TM can be incorporated into a stable, membrane-mimetic thin film over a wide range of surface concentrations by a process of lipid/protein self-assembly and *in situ* photopolymerization. See AA, AB, AC. However, as an alternate strategy using genetically directed synthesis, we have created a short TM construct containing the catalytic region of EGF domains 4-6 along with an artificial amino acid analog, azido (N3)-alanine, incorporated biosynthetically at the C-terminus of the protein (rTM-N3). Through Staudinger ligation with a suitable phosphine PEO derivative (MW 3000), we have generated TM-PEO conjugates. The azidoTM construct and conjugate have been fully characterized by Western blotting and SDS-PAGE. The catalytic activity (k<sub>cat</sub>, K<sub>m</sub>) of the conjugate is comparable to the rTM-N3 mutant alone, as well as a commercially available soluble human TM protein (Solulin™). Of note, by using a biotin or diene terminated PEG derivative, for example, TM can then be rapidly and directly coupled to a surface, which provides an additional approach for linking TM to membrane assemblies.

[00079] Staudinger ligation of azidoTM with phosphine-PEO is not without limitations. The requisite phosphines are susceptible to air oxidation, and their

optimization for improved water solubility and increased reaction rate is synthetically challenging. Thus, we have investigated alternate strategies for direct single-step bioorthogonal coupling of rTM-N3 to target surfaces. Specifically, we have demonstrated that rTM-N3 can be coupled to surfaces via an azide-alkyne [3+2] cycloaddition, termed "click" chemistry (AF, AG). Significantly, both this approach and Staudinger ligation provide a means to selectively conjugate complex biomolecules in richly functionalized environs under mild conditions that do not alter biomolecular activity. The chemical conjugation scheme is summarized in Figure 6. See (AF).

[00080] The rTM-N3 was expressed with an N-terminal S-tag, which allowed the effectiveness of surface coupling to be evaluated using a FITC-labeled anti-S-tag antibody. Likewise, the feasibility of using this strategy to link N3-derivatized carbohydrates to target surfaces was also evaluated using azidolactose, as a model oligosaccharide for more complex polysaccharides, such as heparin. A FITC-labeled lectin was used to ascertain the extent and homogeneity of carbohydrate surface conjugation. This scheme proved quite versatile for surface coupling of both rTM and carbohydrates.

[00081] References for Example 2:

AA. Cazalis CS, Haller CA, Chaikof EL. Site-specific pegylation of a truncated thrombomodulin derivative. *Polymer Mater Sci Eng* 2004;227:198-9.

AB. Cazalis CS, Haller CA, Sease-Cargo L, Chaikof EL. C-terminal site-specific pegylation of a truncated thrombomodulin mutant with retention of full bioactivity. *Bioconjugate Chem* 2004;15:1005-9.

AC. Cazalis CS, Haller CA, Sease-Cargo L, Chaikof EL. Thrombomodulin conjugates. USA. PCT 60/546,436, February 20, 2004.

AD. Kishida A, Akatsuka Y, Yanagi M, Aikou T, Maruyama I, Akashi M. In vivo and ex vivo evaluation of the antithrombogenicity of human thrombomodulin immobilized biomaterials. *ASAIO Journal* 1995;41(3):M369-74.

AE. Vasilets VN, Hermel G, Konig U, Werner C, Muller M, Simon F, et al. Microwave CO<sub>2</sub> plasma-initiated vapour phase graft polymerization of acrylic acid onto polytetrafluoroethylene for immobilization of human thrombomodulin. *Biomaterials* 1997;18(17):1139-45.

- 5 AF. Wang Q, Chan TR, Hilgraf R, Fokin VV, Sharpless KB, Finn MG. Bioconjugation by Copper(I)-Catalyzed Azide-Alkyne [3 + 2] Cycloaddition. *J Am Chem Soc* 2003;125(11):3192-3193.

- AG. Agard NJ, Prescher JA, Bertozzi CR. A strain-promoted [3 + 2] azide-alkyne cycloaddition for covalent modification of biomolecules in living systems. *J Am Chem Soc* 2004;126(46):15046-15047.
- 10

[00082] STATEMENTS REGARDING INCORPORATION BY REFERENCE AND VARIATIONS

- [00083] All references throughout this application, for example patent documents including issued or granted patents or equivalents; patent application publications; and non-patent literature documents or other source material; are hereby incorporated by reference herein in their entireties, as though individually incorporated by reference, to the extent each reference is at least partially not inconsistent with the disclosure in this application (for example, a reference that is partially inconsistent is incorporated by reference except for the partially inconsistent portion of the reference).
- 15
- 20

[00084] Any appendix or appendices hereto are incorporated by reference as part of the specification and/or drawings.

- [00085] Compositions including compounds of the invention can have prodrug forms. Prodrugs of the compounds of the invention are useful in the methods of this invention. Any compound that will be converted in vivo to provide a biologically, pharmaceutically or therapeutically active form of a compound of the invention is a prodrug. Various examples and forms of prodrugs are well known in the art. A biomolecule such as a precursor protein or precursor nucleic acid can be a prodrug. Examples of prodrugs are found, inter alia, in *Design of Prodrugs*, edited by H. Bundgaard, (Elsevier, 1985), *Methods in Enzymology*, Vol. 42, at pp. 309-396, edited
- 25
- 30

by K. Widder, et. al. (Academic Press, 1985); A Textbook of Drug Design and Development, edited by Krosgaard-Larsen and H. Bundgaard, Chapter 5, "Design and Application of Prodrugs," by H. Bundgaard, at pp. 113-191, 1991); H. Bundgaard, Advanced Drug Delivery Reviews, Vol. 8, p. 1-38 (1992); H. Bundgaard, et al., Journal of Pharmaceutical Sciences, Vol. 77, p. 285 (1988); and Nogrady (1985) Medicinal Chemistry A Biochemical Approach, Oxford University Press, New York, pages 388-392).

[00086] Where the terms "comprise", "comprises", "comprised", or "comprising" are used herein, they are to be interpreted as specifying the presence of the stated features, integers, steps, or components referred to, but not to preclude the presence or addition of one or more other feature, integer, step, component, or group thereof. Separate embodiments of the invention are also intended to be encompassed wherein the terms "comprising" or "comprise(s)" or "comprised" are optionally replaced with the terms, analogous in grammar, e.g.; "consisting/consist(s)" or "consisting essentially of/consist(s) essentially of" to thereby describe further embodiments that are not necessarily coextensive.

[00087] The invention has been described with reference to various specific and preferred embodiments and techniques. However, it should be understood that many variations and modifications may be made while remaining within the spirit and scope of the invention. It will be apparent to one of ordinary skill in the art that compositions, methods, devices, device elements, materials, procedures and techniques other than those specifically described herein can be applied to the practice of the invention as broadly disclosed herein without resort to undue experimentation. All art-known functional equivalents of compositions, methods, devices, device elements, materials, procedures and techniques described herein are intended to be encompassed by this invention. Whenever a range is disclosed, all subranges and individual values are intended to be encompassed. This invention is not to be limited by the embodiments disclosed, including any shown in the drawings or exemplified in the specification, which are given by way of example or illustration and not of limitation. The scope of the invention shall be limited only by the claims.

## [00088] REFERENCES

[00089] Wood MJ et al., NMR Structures Reveal How Oxidation Inactivates Thrombomodulin, *Biochemistry* 2003, 42, 11932-11942.

[00090] US 5,126,140, Thrombomodulin-coated biocompatible substance;  
5 5,834,028, Soluble thrombomodulin-containing composition; 5,863,760, Protease-resistant thrombomodulin analogs; 5,583,102, Human thrombomodulin in wound healing; 6,632,791, Thrombomodulin analogs for pharmaceutical use; 5,256,770, Oxidation resistant thrombomodulin analogs; 5,108,759, Endothelial envelopment drug carriers; 6,410,057, Biodegradable mixed polymeric micelles for  
10 drug delivery.

[00091] Weiler, H., Isermann, B. H. Thrombomodulin. (2003) *J. Thrombosis and Haemostasis* 1, 1515-1524.

[00092] Kalafatis, M., Egan, J.O., Van't Veer, C., Cawthern, K. M., Mann, K. G. (1997) The regulation of clotting factors. *Crit. Rev. Eukaryot. Gene Expr.* 7, 241-280.

15 [00093] Esmon, N. L., Owen, W.G., Esmon, C. T. (1982) Isolation of a membrane-bound cofactor for thrombin-catalyzed activation of protein-C. *J. Biol. Chem.* 257, 859-864.

[00094] Esmon, C. T. (1992) The regulation of clotting factors. *Arteriosclerosis and Thrombosis* 12, 135-145.

20 [00095] Parkinson, J. F., Nagashima, M. M, Kuhn, I., Leonard, J., Morser, J. (1992) Structure-Function studies of the epidermal growth-factor domains of human thrombomodulin. *Biochem Biophys. Res. Commun.* 185, 567-576.

[00096] Clarke, J. H., Light, D. R., Blasko, E. J., Parkinson, F., Nagashima, M., McLean, K., Vilander, L., Andrews, W. H., Morser, J., Glaser, C. B. (1993) The short  
25 loop between epidermal growth factor-like domain-4 and domain-5 is critical for human thrombomodulin function. *J. Biol. Chem.* 268, 6309-6315.

[00097] Nagashima, M., Lundh, E., Leonard, J. C., Morser, J., Parkinson J. F. (1993) Alanine-scanning mutagenesis of the epidermal growth factor-like domains of



human thrombomodulin identifies critical residues for its cofactor activity. J. Biol. Chem. 268, 2888-2892.

[00098] Glaser, C. B., Morser, J., Clarke, J. H., blasko, E., McLean, K., Kuhn, I., Chang, R. -J., Lin, J.-H., Vilander, L., Andrews, W. H., and Light, D. R. (1992)

- 5 Oxidation of a secific methionine in thrombomodulin by activated neutrophil products blocks cofactor activity-a potential rapid mechanism for modulation of coagulation. Clin. Invest. 90, 2565-2573.

[00099] Kiick, K. L., Saxon, E., Tirrell, D. A., Bertozzi, C.R. (2002) Incorporation of azides into recombinant proteins for chemoselective modification by the Staudinger  
10 ligation. PNAS 99, 19-24.

[000100] Saxon, E., Bertozzi, C. R. (2000) Cell surface engineering by a modified Staudinger reaction. Science 287, 2007-2010.

[000101] Veronese, F. M. (2001) Peptide and protein PEGylation: a review of problems and solutions. Biomaterials 22, 405-417.

- 15 [000102] Roberts, M. J., Bentley, M. D., Harris, J. M. (2002) Chemistry for peptide and protein PEGylation. Adv Drug delivery reviews 54, 459-476.

[000103] Katre, N. V. (1993) The conjugation of proteins with polyethylene glycol and other polymers. Altering properties of proteins to enhance their therapeutic potential. Adv Drug Delivery Rev 10, 91-114.

- 20 [000104] Monkarsh, S. P., Ma, Y. M., Aglione, A., Bailon, P., Ciolek, D., DeBarbieri, B., Graves, M. C., Hollfelder, K., Michel, H., Palleroni, A., Porter, J. E., Russoman, E., Roy, S., Pan, Y. C. E. (1997) Positional isomers of monopegylated interferon 2a: isolation, characterization, and biological activity. Anal. Biochem. 247, 434-440.

- [000105] Han, H. -S., Yang, S. -L., Yeh, H. -Y., Lin, J. -C., Wu, H. -L., Shi, G. -Y.  
25 (2001) Studies of a novel human thrombomodulin immobilized substrate: surface characterization and anticoagulation activity evaluation. J. Biomater. Sci. Polymer edn, 12, 1075-1089.

- [000106] Tsutsumi, Y., Onda, M., Nagata, S., Lee, B., Kreitman, R. J., Pastan, I. (2000) Site-specific chemical modification with polyethylene glycol of recombinant immunotoxin anti-Tac(Fv)-PE38 (LMB-2) improves antitumor activity and reduces animal toxicity and immunogenicity. *Proc. Natl. Acad. Sci. USA* 97, 8548-8553.
- 5 [000107] He, X. H., Shaw, P. C., Xu, L. H., Tam, S. C. (1999) Site-directed polyethylene glycol modification of trichosanthin: effects on its biological activities, pharmacokinetics, and antigenicity. *Life Sci.* 64, 1163-1175.
- [000108] Goodson, R. J., Katre, N. V. (1990) Site-directed pegylation of recombinant interleukin-2 at its glycosylation site. *Biotechnology* 8, 343-346.
- 10 [000109] Kogan, T.P. (1992) The synthesis of substituted methoxy-poly(ethylene glycol) derivatives suitable for selective protein modification. *Synth. Commun.* 22, 2417-2424.
- [000110] Morpurgo, M., Veronese, F. M., Kachensky, D., Harris, J. M. (1996) Preparation and characterization of poly(ethylene glycol) vinyl sulfone. *Bioconjug. Chem.* 7, 363-368.
- 15 [000111] Woghiren, C., Sharma, B., Stein, S. (1993) Protected thiol-polyethylene glycol: a new activated polymer for reversible protein modification. *Bioconjug. Chem.* 4, 314-318.
- [000112] Karpusas, M., Nolte, M., Benton, C. B., Meier, W., Lipscomb, W. N., Goelz, S. (1997) The crystal structure of human interferon beta at 2.2-A resolution. *Proc. Natl. Acad. Sci. USA* 94, 11813-11818.
- 20 [000113] Kuan, C. T., Wang, Q. C., Pastan, I. (1994) *Pseudomonas* exotoxin A mutants. Replacement of surface-exposed residues in domain II with cysteine residues that can be modified with polyethylene glycol in a site-specific manner. *J. Biol. Chem.* 269, 7610-7616.
- 25 [000114] Yamamoto, Y., Tsutsumi, Y., Yoshioka, Y., Nishibata, T., Kobayashi, K., Okamoto, T., Mukai, Y., Shimizu, T., Nakagawa, S., Nagata, S., Mayumi, T. (2003) Site-specific PEGylation of a lysine-deficient TNF-alpha with full bioactivity. *Nature Biotechnology* 21, 546-552.

[000115] Kurfurst, M. M. (1992) Detection and molecular weight determination of polyethylene glycol-modified hirudin by staining after sodium dodecyl sulfate-polyacrylamide gel electrophoresis. *Anal. Biochem.* 200, 244-248.

5 [000116] Manjula, B. N., Tsai, A., Upadhy, R., Perumalsamy, K., Smith, P. K., Malavalli, A., Vandegriff, K., Winslow, R. M., Intaglietta, M., Prabhakaran, M., Friedman, J. M., Acharya, A. S. (2003) Site-specific PEGylation of hemoglobin at cys-93(beta): Correlation between the colligative properties of the PEGylated protein and the length of the conjugated PEG chain. *Bioconjugate Chemistry* 14, 464-472.

10 [000117] Feng, J., Tseng, P. -Y., Faucher, K. M., Orban, J. M., Sun, X. -L., Chaikof, E. L. (2002) Functional reconstitution of thrombomodulin within a substrate-supported membrane-mimetic polymer film. *Langmuir* 18, 9907-9913.

[000118] Kishida, A., Ueno, Y., Maruyama, I., Akashi, M. (1994) Immobilization of human thrombomodulin on biomaterials-Evaluation of the activity of immobilized human thrombomodulin. *Biomaterials* 15, 1170-1174.

15 [000119] Kishida, A., Ueno, Y., Maruyama, I., Akashi, M. (1994) Immobilization of human thrombomodulin on onto poly(ether urethane urea) for developing antithrombogenic blood-contacting materials. *Biomaterials* 15, 848-852.